

具有重要意义<sup>[10]</sup>，为此，本研究为进一步研究 MC-FA 调节 HDL 的作用提供了依据和奠定了基础。

## 参考文献

[1] Aoyama T, Nosaka N, Kasai M. Research on the nutritional characteristics of medium-chain fatty acids [J]. *J Med Invest*, 2007, 54: 385–388.

[2] Xue CY, Liu YH, Wang J, et al. Consumption of medium- and long-chain triacylglycerols decreases body fat and blood triglyceride in Chinese hypertriglyceridemic subjects [J]. *Euro J Clin Nutr*, 2009, 6: 879–886.

[3] Liu YH, Wang J, Zhang RX, et al. A good response to oil with medium- and long-chain fatty acids in body fat and blood lipid profiles of male hypertriglyceridemic subjects [J]. *Asia Pac J Clin Nutr*, 2009, 18 (3): 351–358.

[4] Ogawa A, Nosaka N, Kasai M, et al. Dietary medium- and long-chain triacylglycerols accelerate diet-induced thermogenesis in humans [J]. *J Oleo Sci*, 2007, 56 (6): 283–287.

[5] Nagao K, Yanagita T. Medium-chain fatty

acids: functional lipids for the prevention and treatment of the metabolic syndrome [J]. *Pharmacol Res*, 2010, 61 (3): 208–212.

[6] Xue CY, Liu YH, Wang J, et al. Chinese hypertriglyceridemic subjects of different ages responded differently to consuming oil with medium- and long-chain fatty acids [J]. *Biosci Biotech Bioch*, 2009, 73 (8): 1711–1717.

[7] Toth PP. High-density lipoprotein as a therapeutic target: clinical evidence and treatment strategies [J]. *Am J Cardiol*, 2005, 96 (9A): 50K–58K.

[8] Tardif JC, Grégoire J, L'Allier PL, et al. Effects of reconstituted high-density lipoprotein infusions on coronary atherosclerosis: a randomized controlled trial [J]. *JAMA*, 2007, 297 (15): 1675–1682.

[9] Shi W, Wang X, Wong J, et al. Effect of macrophage-derived apolipoprotein E on hyperlipidemia and atherosclerosis of LDLR-deficient mice [J]. *Biochem Biophys Res Commun*, 2004, 317 (1): 223–229.

[10] Toth PP. The “Good Cholesterol” High-Density Lipoprotein [J]. *Circulation*, 2005, 111: e89–e91.

## 17 种常见蔬菜不同部位抗氧化活性的比较研究

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**摘要:** 目的 比较蔬菜不同部位的抗氧化活性。方法 采用 FRAP 法、2, 4-二硝基苯肼法、Folin-Ciocalteu 法、铝离子显色法分别测定 17 种常见蔬菜不同部位的 FRAP 值及 VC、总多酚、总黄酮的含量, 并分析了总抗氧化活力与抗氧化物质含量之间的相关性。结果 蔬菜叶子、皮的抗氧化活性高于其对应的茎和肉质部分; 藕皮的总抗氧化活性最高, 其次为藕肉、茴香叶、香菜叶、姜肉等, 黄瓜肉的总抗氧化活性最弱; 所有水提取物的总抗氧化活力均高于丙酮提取物; 水提取物中, 蔬菜抗氧化活性与 VC、多酚、黄酮含量的相关性具有统计学意义, 多元回归分析结果显示多酚对蔬菜总抗氧化活力的贡献最大。结论 蔬菜不同部位的抗氧化活性差异较大, 多酚类物质在蔬菜体外抗氧化活性方面可能起主要作用。

**关键词:** 蔬菜; 抗氧化活性; FRAP 值; VC; 总多酚; 总黄酮

过多的活性氧、活性氮自由基会引起机体内的氧化应激,并导致碳水化合物、脂类、蛋白质及核酸的氧化损伤。据报道,氧化应激会导致人类很多疾病的发生,如心血管疾病、肿瘤等<sup>[1-5]</sup>。膳食抗氧化剂被认为能够有效地保护氧化应激引起的损伤<sup>[6-8]</sup>。蔬菜水果中富含维生素、矿物质、纤维素,同时还含有天然的抗氧化物质,如类胡萝卜素、多酚、黄酮等。这些抗氧化物质能够有效地清除多种自由基、抑制链式反应的起始或者通过结合金属离子减少自由基的产生<sup>[9-12]</sup>。大量流行病学资料显示,蔬菜水果的摄入量与一些慢性疾病的发生之间呈现显著的负相关<sup>[13-16]</sup>。因此,在预防氧化应激相关疾病方面<sup>[17-19]</sup>,增加蔬菜水果的摄入量常常成为推荐的策略之一。

我们实验室之前通过测定了水果皮、果肉、种子部位的抗氧化活性,发现水果皮及种子的抗氧化活性高于果肉部分,表明水果果皮、种子部分可能是较有价值的天然抗氧化剂来源<sup>[20]</sup>。国外也有类似的报道<sup>[21,22]</sup>。然而,蔬菜不同部位抗氧化活性的研究鲜有报道<sup>[23]</sup>。在我们日常生活中,蔬菜一些部位因为卫生问题、口感不好、烹饪问题常常被丢弃,因此,可能导致蔬菜中潜在抗氧化物质及其它营养素的丢失。

本次实验中,我们采用 FRAP 法测定蔬菜不同部位抗氧化活性,并分析了其与抗氧化物质 VC、多酚、黄酮含量的相关性。

## 1 材料与方法

### 1.1 材料与处理

藕、茴香、姜、香菜、菠菜、油菜、小白菜、茄子、芹菜、生菜、土豆、大白菜、青椒、韭菜、卷心菜、花菜、黄瓜、大葱共 17 种,购自天津市农贸市场。所有样品用自来水、蒸馏水反复冲洗干净,晾干表面水分,按不同部位,称取 1~5g,在研钵内按 1:10 比例加入蒸馏水,冰浴研磨制备匀浆液,3000r/min 离心,15min 后取上清液,残渣重复提取两次,将三次上清液混合备用。水溶液提取之后,向残渣中按比例加入适量丙酮提取,室温 3000r/min 离心 15min,重复一次,将两次上清液混合备用。水溶性及脂溶性上清液用于测定蔬菜的水溶性及脂溶性抗氧化活性、VC、总多酚、水溶性及脂溶性总黄酮含量,每份样品重复测定 3 次。

### 1.2 方法

#### 1.2.1 抗氧化活性

采用 FRAP 法测定。参照 Benzie 与 Strain 的方

法<sup>[24]</sup>,并适当的优化。原理:在低 pH 值下,样品中的还原物质将  $Fe^{3+}$ -三吡啶三吡嗪 (tripipyridy-triazine,  $Fe^{3+}$ -TPTZ) 还原为二价铁形式,呈现出明显的蓝色,于 593nm 处有最大光吸收。根据吸光度大小计算样品抗氧化活性的强弱。TPTZ、Rutin, 购自 Sigma 公司;其余均为国产试剂,分析纯级。

#### 1.2.2 VC

采用 2,4-二硝基苯肼法<sup>[25]</sup>,以抗坏血酸为标准。原理:还原型抗坏血酸被活性炭将氧化为脱氢抗坏血酸,然后与 2,4-二硝基苯肼在 37℃ 水浴 3h 作用生成红色的脎,在浓硫酸的脱水作用下,可转变为橘红色的化合物-双-2,4-二硝基苯,在硫酸溶液中显色稳定,吸光度与总抗坏血酸含量成正比,一般根据 490nm 下的吸光度大小计算样品 VC 含量。

#### 1.2.3 总多酚

采用 Folin-Ciocalteus 法<sup>[26]</sup>,以没食子酸为标准。原理为在碱性溶液中,酚类化合物可将钨钼酸还原 ( $W_6^{+}$  变为  $W_5^{+}$ ),生成蓝色化合物,颜色的深浅与酚含量成正相关,在 760 nm 处有最大光吸收。

#### 1.2.4 总黄酮

采用铝离子显色法<sup>[27]</sup>,以芦丁 (Rutin, Sigma) 为标准。原理:黄酮类化合物是一类具有苯并吡喃环结构的化合物,其 3-羟基、4-羟基或 5-羟基、4-羰基及邻二位酚羟基,与铝盐进行络合反应,在碱性条件下生成红色的络合物,用于 510nm 测定其吸光度,与芦丁标准品比较进行定量。

## 1.3 数据处理

结果用均值 ± 标准差表示,所有数据用 SPSS10.0 软件进行统计分析。

## 2 结果与讨论

### 2.1 蔬菜不同部位的抗氧化活性

17 种常见蔬菜不同部位结果见表 1。在测定的所有蔬菜中,藕皮的总抗氧化活力最高,达到 19.45mmol/100g;藕肉、茴香叶、香菜叶、菠菜叶、青椒肉、生姜皮、油菜叶、小白菜叶、生姜肉、芹菜叶、茄子皮、生菜叶、大葱叶、土豆皮次之,总抗氧化活力值在 3.06 到 1.11 mmol/100g;青椒肉、大白菜叶、青椒籽、卷心菜叶、花菜花、大葱茎、黄瓜皮、土豆肉、花菜茎、茴香茎、香菜茎、小白菜茎、油菜茎、卷心菜茎、茄子肉、菠菜茎、大白菜茎、生菜茎、芹菜茎、黄瓜肉等抗氧化活力值相对较低,在 0.87 mmol/100g 以下;抗氧化活力值最强与最弱的相差 216 倍。

Table 1 FRAP values of different fractions of 17 vegetables (mmol/100g wet weight)

Vegetables	Fraction		Total	Rank
	Water	Acetone		
Bok choy leaf	0.81 ± 0.08	0.04 ± 0.00	0.85	16
Bok choy stem	0.16 ± 0.01	—	0.16	27
Cabbage leaf	0.58 ± 0.04	0.04 ± 0.00	0.63	18
Cabbage stem	0.21 ± 0.02	0.02 ± 0.00	0.24	25
Cauliflower flower	0.46 ± 0.01	0.03 ± 0.00	0.5	19
Cauliflower stem	0.39 ± 0.03	0.03 ± 0.00	0.42	21
Celery leaf	1.02 ± 0.11	0.86 ± 0.03	1.88	10
Celery stem	0.10 ± 0.02	—	0.1	29
Coriander leaf	2.09 ± 0.14	0.80 ± 0.03	2.89	4
Coriander stem	0.23 ± 0.02	0.05 ± 0.00	0.28	23
Cucumber peel	0.35 ± 0.02	0.12 ± 0.00	0.47	20
Cucumber pulp	0.09 ± 0.01	—	0.09	30
Eggplant peel	1.30 ± 0.09	0.03 ± 0.00	1.33	11
Eggplant pulp	0.24 ± 0.02	—	0.24	25
Endive lettuce leaf	0.85 ± 0.04	0.44 ± 0.03	1.28	12
Endive lettuce stem	0.14 ± 0.00	—	0.14	28
Fennel leaf	2.39 ± 0.15	0.59 ± 0.04	2.98	3
Fennel stem	0.26 ± 0.02	0.14 ± 0.01	0.4	22
Ginger peel	1.91 ± 0.06	0.57 ± 0.04	2.48	6
Ginger pulp	1.71 ± 0.08	0.24 ± 0.05	1.95	9
Green pepper pulp	0.82 ± 0.01	0.05 ± 0.01	0.87	15
Green pepper seed	0.73 ± 0.02	0.08 ± 0.01	0.81	17
Lotus root peel	17.56 ± 1.18	1.89 ± 0.12	19.45	1
Lotus root pulp	2.90 ± 0.10	0.16 ± 0.01	3.06	2
Pak choy leaf	1.63 ± 0.13	0.32 ± 0.03	1.96	8
Pak choy stem	0.26 ± 0.00	—	0.26	24
Potato peel	0.82 ± 0.03	0.29 ± 0.01	1.11	14
Potato pulp	0.42 ± 0.01	—	0.42	21
Rape leaf	1.64 ± 0.08	0.56 ± 0.05	2.2	7
Rape stem	0.21 ± 0.01	0.02 ± 0.00	0.24	25
Scallion leaf	1.02 ± 0.05	0.15 ± 0.02	1.17	13
Scallion stem	0.48 ± 0.05	0.02 ± 0.00	0.5	19
Spinach leaf	1.67 ± 0.06	0.87 ± 0.07	2.54	5
Spinach stem	0.12 ± 0.00	0.07 ± 0.01	0.19	26

Data are expressed as mean ± SD. Each vegetable fraction was analyzed three times.

“—”, not detectable.

所有水提取物的抗氧化值高于丙酮提取物。其中大白菜茎、芹菜茎、黄瓜肉、茄子肉、生菜茎、小白菜茎及土豆肉未测得丙酮提取物的抗氧化活力值。实验结果显示,FRAP 法测得所有蔬菜的皮及叶部分高

于其对应的肉及茎部分的抗氧化值,说明皮及叶子部分富含抗氧化物质,或者含有的物质具有较强的抗氧化活性。

## 2.2 蔬菜不同部位 VC、多酚、黄酮含量

蔬菜中含有大量的化合物发挥着抗氧化作用,如抗坏血酸、 $\beta$ -胡萝卜素、多酚、黄酮及其他物质<sup>[28-30]</sup>。本实验测定了17种蔬菜不同部位抗坏血酸、总多酚及黄酮的含量(见表2)。本实验结果显示,芹菜叶、藕皮及茴香叶抗坏血酸含量高于200 mg/100g;香菜叶、青椒肉、花菜花、油菜叶、花菜茎、小白菜茎及卷心菜茎抗坏血酸含量次之,在156.55 to 103.01mg/100g;余下蔬菜不同部位的抗坏血酸含量低于100mg/100g,土豆皮未测出抗坏血酸。对实验结果进行比较之后发现,除了土豆之外,所有蔬菜皮及叶部分抗坏血酸浓度高于其对应的肉及茎部位。

在17种蔬菜中,藕皮含的多酚最高,达到228.10

mg/100g;多酚含量超过100 mg/100g的有茴香叶、土豆皮、香菜叶;剩余蔬菜各个部位的多酚含量低于100 mg/100g。同样地,蔬菜皮及叶子部位多酚含量高于其对应的果肉及茎。Strack 已证实多酚在紫外线辐射、病原体及外来者入侵方面发挥着重要的保护作用<sup>[31]</sup>。

水提取物中,总黄酮含量最高的是藕皮;生姜皮、茴香叶、香菜叶、生姜肉等次之;卷心菜叶、芹菜叶和茎、黄瓜皮及果肉、茄子肉、生菜茎、土豆皮、油菜茎、大白菜叶和茎未测出水溶性黄酮含量。丙酮提取物中,青椒籽总黄酮含量最高;藕皮、生姜皮、土豆皮、花菜花、生姜肉、花菜茎、藕肉、土豆肉次之;剩余的蔬菜未测出脂溶性黄酮含量。

Table 2 The contents of VC, phenolics and flavonoids of different fractions of vegetables

Vegetables	VC (mg/100gFW)	Total phenolics (mgGA/100gFW)	Flavonoids (mgRutin/100gFW)		
			Water	Acetone	Total
Bok choy leaf	36.91 ± 0.39	49.41 ± 1.33	-	-	-
Bok choy stem	8.50 ± 1.00	2.03 ± 0.15	-	-	-
Cabbage leaf	103.01 ± 7.22	37.08 ± 1.60	3.64 ± 0.77	-	3.64
Cabbage stem	21.96 ± 1.46	12.02 ± 1.91	-	-	-
Cauliflower flower	123.20 ± 1.00	52.46 ± 1.17	35.87 ± 3.52	14.63 ± 2.97	50.5
Cauliflower stem	117.27 ± 3.99	22.97 ± 2.70	22.09 ± 1.54	7.07 ± 1.12	29.16
Celery leaf	247.69 ± 9.57	77.10 ± 7.35	-	-	-
Celery stem	21.70 ± 0.77	14.16 ± 1.41	-	-	-
Coriander leaf	156.55 ± 9.52	108.38 ± 10.21	93.42 ± 1.54	-	93.42
Coriander stem	60.61 ± 2.25	19.97 ± 1.04	10.67 ± 1.89	-	10.67
Cucumber peel	61.63 ± 5.23	43.08 ± 4.04	-	-	-
Cucumber pulp	27.11 ± 3.57	9.92 ± 0.89	-	-	-
Eggplant peel	51.88 ± 2.52	98.64 ± 8.99	11.64 ± 1.78	-	11.64
Eggplant pulp	12.93 ± 0.93	24.28 ± 2.12	-	-	-
Endive lettuce leaf	89.20 ± 12.97	98.64 ± 1.54	8.44 ± 0.96	-	8.44
Endive lettuce stem	3.69 ± 0.68	5.53 ± 0.52	-	-	-
Fennel leaf	204.00 ± 17.60	110.15 ± 3.11	98.76 ± 8.87	-	98.76
Fennel stem	27.37 ± 2.77	17.59 ± 1.17	8.98 ± 0.77	-	8.98
Ginger peel	23.36 ± 1.93	91.96 ± 0.77	99.87 ± 8.46	55.61 ± 4.18	155.48
Ginger pulp	15.17 ± 1.94	72.46 ± 1.17	91.78 ± 6.64	14.05 ± 1.19	115.83
Green pepper pulp	124.53 ± 9.92	40.44 ± 3.43	22.98 ± 3.35	-	22.98
Green pepper seed	26.97 ± 1.08	53.77 ± 1.94	34.56 ± 2.54	231.79 ± 13.17	266.35
Lotus root peel	241.93 ± 29.89	228.10 ± 4.80	147.64 ± 9.64	77.70 ± 7.08	225.34
Lotus root pulp	95.36 ± 6.38	60.50 ± 6.91	62.98 ± 6.11	6.32 ± 0.63	69.3
Pak choi leaf	105.20 ± 2.27	71.44 ± 9.18	70.76 ± 6.24	-	70.76
Pak choi stem	25.50 ± 4.50	16.12 ± 1.89	6.77 ± 1.02	-	6.77
Potato peel	-	108.50 ± 3.91	-	53.70 ± 1.49	53.7
Potato pulp	22.84 ± 2.12	73.10 ± 7.21	12.33 ± 1.89	0.76 ± 0.01	13.09
Rape leaf	121.91 ± 3.04	78.61 ± 2.47	80.98 ± 8.58	-	80.98
Rape stem	31.10 ± 2.16	6.55 ± 0.68	-	-	-
Scallion leaf	86.54 ± 2.30	91.12 ± 1.78	21.20 ± 2.81	-	21.2
Scallion stem	18.58 ± 0.71	51.42 ± 0.51	14.03 ± 2.10	-	14.03
Spinach leaf	62.94 ± 1.00	88.90 ± 8.5	83.64 ± 6.58	-	83.64
Spinach stem	14.95 ± 2.87	8.60 ± 1.6	4.17 ± 0.51	-	4.17

Data are expressed as mean ± SD. Each vegetable fraction was analyzed three times.

"-", not detectable.

### 2.3 蔬菜不同部位的 FRAP 值与抗坏血酸、总多酚及黄酮含量的相关性分析

在水提取物中,蔬菜不同部位的 FRAP 值与抗坏血酸、多酚、黄酮含量之间呈显著性正相关 ( $R^2 = 0.2983, 0.4761, 0.4984$ );在丙酮提取物中,FRAP 值与黄酮含量之间的相关性不具有统计学意义 ( $R^2 =$

0.0341)。对本实验数据进行多元回归分析,结果显示多酚在不同部位蔬菜抗氧化力方面发挥主要作用(表3)。我们的研究结果与 Maisuthisakul 等、杨冬梅等、陆广念等的研究一致,进一步证实蔬菜的抗氧化活性与酚类物质直接相关<sup>[32-36]</sup>。

Table 3 Results of multivariate regression analysis

Variable	$\beta$	S. E	Beta	t	P
constant	-1.739	0.531	-	-3.275	0.003**
VC	7.271E-03	0.006	0.148	1.124	0.270
Phenolics	4.349E-02	0.011	0.599	3.795	0.001**
Flavonoids	9.912E-02	0.006	0.201	1.575	0.126

Significant difference, \*\*  $P < 0.05$ .

## 3 结论

### 3.1 蔬菜不同部位的抗氧化活性差异很大

蔬菜皮、叶子的抗氧化活性一般高于茎和肉质,水提取物的抗氧化活性高于丙酮提取物。

### 3.2 蔬菜不同部位的 VC、多酚、黄酮含量差异很大

其中多酚类物质是蔬菜体外抗氧化活性的主要来源。

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## 参考文献

[1] Temple, N. J. Antioxidants and disease: More questions than answers[J]. Nutrition Research, 2000, 20:449-459.

[2] Moskovitz, J., Yim, K. A., & Choke, P. B. Free radicals and disease[J]. Archives of Biochemistry and Biophysics, 2002, 397:354-359.

[3] World Health Organization. Report of a Joint FAO/WHO Expert Consultation: Diet, Nutrition and the Prevention of Chronic Disease. Technical Report Series no. 916. 2003, Geneva: WHO.

[4] Atoui, A. K., Mansouri, A., Boskou, G., & Kefalas, P. Tea and herbal infusions: their antioxidant activity and phenolic profile[J]. Food Chemistry, 2005, 89:27-36.

[5] Scalbert, A., Manach, C., Morand, C., Reme-

sy, C. & Jimenez, L. Dietary polyphenols and the prevention of diseases[J]. Critical Reviews in Food Science and Nutrition, 2005, 45:287-306.

[6] Wang, L. S., & Stoner, G. D. Anthocyanins and their role in cancer prevention[J]. Cancer Letters, 2008, 269:281-290.

[7] Huxley, R. R., & Neil, H. A. W. The relationship between dietary flavonol intake and coronary heart disease mortality: A meta-analysis of prospective cohort studies[J]. European Journal of Clinical Nutrition, 2003, 57(8):904-908.

[8] Knekt, P., Kumpulainen, J., Jarvinen, R., Rissanen, H., Heliövaara, M., Rissanen, A., et al. Flavonoid intake and the risk of chronic diseases. American Journal of Clinical Nutrition, 2002, 76:560-568.

[9] Kris-Etherton, P. M., Hecker, K. D., Bonanome, A., Coval, S. M., Binkoski, A. E., Hilpert, K. F. Bioactive compounds in foods: their role in the prevention of cardiovascular disease and cancer[J]. American Journal of Medicine, 2002, 30:71-88.

[10] Peschel, W., Sanchez-Rabaneda, F., Diekmann, W., Plescher, A., Gartzia, I., Jimenez, D., et al. An industrial approach in the search of natural antioxidants from vegetable and fruit wastes[J]. Food Chemistry, 2006, 97:137-150.

[11] Ohr, L. M. Dietary antioxidants[J]. Food Technology, 2004, 58:67-74.

[12] Barreira, C. M. J., Ferreira, C. F. R. I., Beatriz, M., Oliveira, P. P. J., & Pereira, A. Antioxidant activities of the extracts from chestnut flower, leaf,

- skins and fruit[J]. *Food Chemistry*,2008,107:1106 – 1113.
- [13] Dauchet, L. , Amouyel, P. , Herchberg, S. , et al. . Fruit and vegetable consumption and risk of coronary heart disease;a meta-analysis of cohort studies[J]. *Nutrition*,2006,136:2588 – 2593.
- [14] Gossiau, A. , Chen, K. Y. . Nutraceuticals, apoptosis, and disease prevention[J]. *Nutrition*,2004,20: 95 – 102.
- [15] Gundgaard, J. N. , Nielson, J. N. , Olsen, J. , & Sorensen, J. . Increased intake of fruit and vegetables: Estimation of impact in terms of life expectancy and healthcare costs[J]. *Public Health Nutrition*,2003,6:25 – 30.
- [16] Johnsen, S. , Overvad, K. , Stripp, C. , Tjonne-land, A. , Husted, S. E. , & Sorensen, H. T. . Intake of fruit and vegetables and the risk of ischemic stroke in a cohort of Danish men and women. *American Journal of Clinical of nutrition*,2003,78:57 – 64.
- [17] Kaliora, A. C. , Dedoussis, G. V. Z. , & Schmidt, H. . Dietary antioxidants in preventing atherosclerosis[J]. *Atherosclerosis*,2006,187:1 – 17.
- [18] Ramassamy, C. . Emerging role of polyphenolic compounds in the treatment of neurodegenerative diseases:A review of their intracellular targets[J]. *European Journal of Pharmacology*,2006,545:51 – 64.
- [19] Pietta, P. , Minoggio, M. & Bramati, L. . Plant polyphenols: Structure, occurrence and bioactivity [ J ]. *Studies in Natural Products Chemistry*,2003,28:257 – 312.
- [20] Guo, C. J. , Yang, J. J. , Wei, J. Y. , Li, Y. F. , Xu, J. , & Jiang, Y. G. . Antioxidant activities of peel, pulp and seed fractions of common fruits as determined by FRAP assay [ J ]. *Nutrition Research*,2003, 23:1719 – 1726.
- [21] Murthy, K. N. C. , Jayaprakasha, G. K. , & Singh, R. P. . Studies on antioxidant activity of pomegranate peel extract using in vivo models[J]. *Journal of Agricultural and Food Chemistry*,2002,50:4791 – 4795.
- [22] Tomas-Barberan, F. A. , Gil, M. I. , Cremin, P. , Waterhouse, A. L. , Hess-Pierce, B. , & Kader, A. A. . HPLC-DAD-ESIMS analysis of phenolic compounds in nectarines, peaches, and plums[J]. *Journal of Agricultural and Food Chemistry*,2001,49:4748 – 4760.
- [23] Park, J. , Kim, J & Kim, MK. . Onion flesh and onion peel enhance antioxidant status in aged rats [J]. *Journal of nutritional Science and vitaminology*, 2007,53:21 – 29.
- [24] Benzie, I. F. F. & Strain, J. J. . The ferric reducing ability of plasma (FRAP) as a measure of “antioxidant power”: the FRAP assay [J]. *Analytical Biochemistry*,1996,239:70 – 76.
- [25] GB12392 – 90. Method for determination of total ascorbic acid in fruits, vegetables and derived products [ M ]. Approved by Ministry of Public Health, PR China,1990, pp. 388 – 389.
- [26] Singleton, V. L. , Orthofer, R. & LamuelaRaventos, R. M. . Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent [ J ]. *Methods Enzymol*,1999,299:152 – 178.
- [27] Jia, Z. , Tang, M. , & Wu, J. . The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals [ J ]. *Food Chemistry*, 1999,64:555 – 559.
- [28] Heim, K. E. , Tagliaferro, A. R. , & Bobilya, D. J. . Flavonoid antioxidants: Chemistry, metabolism and structure-activity relationships [ J ]. *Journal of Nutritional Biochemistry*,2002,13:572 – 584.
- [29] Seifried, H. E. . Oxidative stress and antioxidants:A link to disease and prevention? [ J ]. *Journal of Nutritional Biochemistry*,2007,18:168 – 171.
- [30] Brat, P. , George, S. , Bellamy, A. , Du Chaufaut, L. , Scalbert, A. , Mennen, L. , et al. . Daily polyphenol intake in France from fruit and vegetables [ J ]. *Journal of Nutrition*,2006,136:2368 – 2373.
- [31] Strack, D. . Phenolic metabolism [ M ]. In P. M. Dey & J. B. Harborne (Eds. ), 1997, *Plant Biochemistry* ( pp. 387 – 416). London :Academic Press.
- [32] Maisuthisakul, P. , Pongsawatmanit, R. & Gordon, M. H. . Characterization of the phytochemicals and antioxidant properties of extracts from teaw ( *Cratogeomys formosum* Dyer) [ J ]. *Food Chemistry*,2007,100:1620 – 1629.
- [33] Abu Bakar, M. F. , Mohamed, M. , Rahmat, A. & Fry, J. . Phytochemicals and antioxidant activity of different parts of bambangan ( *Mangifera pajang*) and tarap ( *Artocarpus odoratissimus*) [ J ]. *Food Chemistry*,2009, 113:479 – 483.
- [34] Kaur, C. & Kapoor, H. C. . Antioxidant activity and total phenolic content of some Asian vegetables [ J ]. *International Journal of Food Science and Technology*,2002,37:153 – 162.

[35]杨冬梅,金月婷等. 12种常见蔬菜抗氧化活性的比较研究[J]. 中国食品学报,2007,7(5):24-29.

[36]陆广念,宋晓敏等. 扬州常见蔬菜的抗氧化活性[J]. 河南工业大学学报:自然科学版,2009,30(5):35-38.

## 氧化应激对 Met 负载后大鼠肝细胞 Hcy 代谢的影响

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**摘要:**目的 研究氧化应激对 Met 负载后大鼠肝细胞 Hcy 及相关氨基酸代谢的影响。方法 体外培养 BRL 大鼠肝细胞,选用 100 $\mu$ mol/L H<sub>2</sub>O<sub>2</sub> 诱导氧化应激,选用 50 mmol/L Met 诱导蛋氨酸负载。将大鼠肝细胞分为对照组、氧化应激组(100 $\mu$ mol/L H<sub>2</sub>O<sub>2</sub> 作用 2h)、Met 负载组(50 mmol/L Met 作用 1h)、氧化应激 + Met 负载组(100 $\mu$ mol/L H<sub>2</sub>O<sub>2</sub> 作用 2h + 50 mmol/L Met 作用 1h),实验结束收集各组培养液上清。采用高效液相法测定 Hcy、Cys 和 GSH 的含量,采用全自动氨基酸分析仪测定相关氨基酸的含量。结果 与对照组比较, Met 负载组 Hcy、Cys、GSH 的含量显著增加( $P < 0.05$ ),氧化应激 + Met 负载组 Hcy、Cys 的含量显著增加( $P < 0.05$ )。与氧化应激组比较, Met 负载组和氧化应激 + Met 组的 Hcy、Cys、GSH 含量均显著增加( $P < 0.05$ )。与对照组比较, Ser、Tau、Glu 和 Gly 的含量在氧化应激组、Met 负载组和氧化应激 + Met 组均显著减少( $P < 0.05$ )。氧化应激组的 Ser 和 Gly 含量显著低于 Met 负载组( $P < 0.05$ ),而 Tau 含量显著高于 Met 负载组( $P < 0.05$ );氧化应激组的 Tau 含量显著高于氧化应激 + Met 负载组( $P < 0.05$ ),而 Gly 含量显著低于氧化应激 + Met 负载组( $P < 0.05$ )。Met 负载组的 Ser、Glu、Gly 显著高于氧化应激 + Met 负载组( $P < 0.05$ ),而 Tau 显著低于氧化应激 + Met 负载组( $P < 0.05$ )。结论 氧化应激对 Met 负载后大鼠肝细胞的 Hcy 代谢可能具有一定的促进作用。

**关键词:**蛋氨酸负载;同型半胱氨酸;氧化应激

## Effects of oxidative stress on Hcy metabolism in Met loading rat hepatocytes

**Abstract: Objective** To investigate effects of oxidative stress on Hcy and related amino acids metabolism. **Methods** 100 $\mu$ mol/L H<sub>2</sub>O<sub>2</sub> was selected to induce oxidative stress, and 50 mmol/L Met was selected to induce Met loading. Cultured BRL rat hepatocytes was divided into control, oxidatively stressed, Met loading, and oxidatively stressed + Met loading groups. At the end of the experiment, culture fluid was collected. Hcy, Cys and GSH were measured by HPLC, amino acids were assayed by Amino Acids Analyzer. **Results** Compared to control, the contents of Hcy, Cys and GSH significantly increased in Met loading group ( $P < 0.05$ ), and the contents of Hcy and Cys also increased in oxidatively stressed + Met loading group ( $P < 0.05$ ). Compared to oxidatively stressed group, the concentrations of Hcy, Cys and GSH distinctively increased in Met loading and oxidatively stressed + Met loading groups ( $P < 0.05$ ). The contents of Ser, Tau, Glu and Gly decreased in oxidatively stressed, Met loading and oxidatively stressed + Met loading groups ( $P < 0.05$ ) compared to control. The concentrations of Ser and Gly in oxidatively stressed group were significantly lower, while Tau was higher than in Met loading group ( $P < 0.05$ ). The content of Tau in oxidatively stressed group was higher while Gly was lower than in oxidatively stressed + Met loading group ( $P < 0.05$ ). The concentrations of Ser, Glu, and Gly in Met loading group were higher, while Tau was lower than in oxida-